iD Buffer *composition*

5x Sample Buffer:

	SDS-PAGE	**Native PAGE**	**Final concentration** (1x Buffer)
Tris base	300 mg	300 mg	50 mM
Glycerol	5.0 mL	5.0 mL	10%
Bromophenol blue	25 mg	25 mg	0.01 %
2-mercaptoethanol	1.0 mL	1.0 mL	2%
SDS	1.0 g		2%
Deinonized water	to 10 mL	to 10 mL	
pH (use 8M NaOH or 8M HCl)	6.8	6.8	

Store at room temperature

1x protein sample solution:

Sample	x µL
Sample Buffer 5x	2 µL
Deinonized water	to 10 µL

Heat at 95°C for 10 minutes before loading on gel

10x MOPS Running Buffer: to separate medium to large proteins >20 kDa

	SDS-PAGE	**Native PAGE**	**Final concentration** (1x Buffer)
Tris base	60.6 g	60.6 g	50 mM
MOPS	104.6 g	104.6 g	50 mM
EDTA	3.0 g	3.0 g	1 mM
SDS	10.0 g		0.1 %
Deinonized water	to 1000 mL	to 1000 mL	
pH (do not adjust)	7.7	7.7	

Store at 4°C up to 6 months

10x MES Running Buffer: to separate small proteins (2-50 kDa)

	SDS-PAGE	**Native PAGE**	**Final concentration** (1x Buffer)
Tris base	60.6 g	60.6 g	50 mM
MES	97.6 g	97.6 g	50 mM
EDTA	3.0 g	3.0 g	1 mM
SDS	10.0 g		0.1 %
Deinonized water	to 1000 mL	to 1000 mL	
pH (adjust with 8M NaOH or 8M HCl)	7.3	7.3	

Store at 4°C up to 6 months

20x Transfer Buffer:

	20x Transfer Buffer*
Tris base	60.6 g
Bicine	81.6 g
Deionized water	1000 mL

To make 1L of 1x transfer buffer: mix 50 ml of 20x transfer buffer, 100 mL of methanol or ethanol and 850 mL of deionized water.

FOR FURTHER INFORMATION PLEASE CONTACT OUR CUSTOMER HELP DESK:

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