

# Takyon® Etheris

## SYBR® 2X qPCR Master Mix Blue<sup>[1]</sup>

**UF-ESMT-B0101**
**UF-ESMT-B0701**
**UF-ESMT-B0705**
**UF-ESMT-B0710**

[1.5 mL]

[7.5 mL]

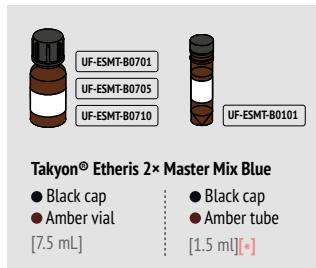
[5 packs of 7.5 mL]

[150 x 20 µL RXNs]

[750 x 20 µL RXNs]

[10 packs of 7.5 mL]

### Kit content (for 750 (150<sup>[\*]</sup>) - 20 µL reactions)

**INCLUDED**

**Takyon® Etheris 2X Master Mix Blue**

- Black cap
- Amber vial

[7.5 mL]

- Black cap
- Amber tube

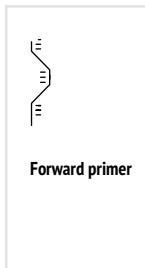
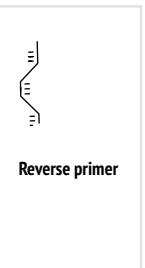
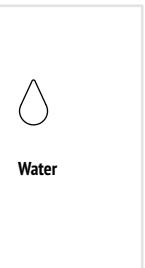
[1.5 mL]<sup>[\*]</sup>

**OPTIONAL**

**ROX™**

- Red cap
- Amber tube

[0.5 mL]

**NOT INCLUDED**

**Forward primer**

**Reverse primer**

**Water**

— Takyon® HotStart DNA polymerase

— SYBR® Green

— Inert Blue Dye

— MgCl<sub>2</sub> (2.5 mM final concentration)

— dNTPs

— Stabilizers

For signal normalization

### Storage

Takyon® kit must be stored in a constant temperature freezer in the dark.

**SHORT TERM STORAGE**

6 Month stability

4°C

**LONG TERM STORAGE**

12 Month stability

 -15°C  
-25°C

### Optimization tips

Refer to the primer design guidelines, custom assay design recommendations and primer titration matrix for best results.

For new users of this kit, conducting a primer titration matrix may be required to ensure optimal performance.


**TECHNICAL  
INFORMATION**

<sup>[1]</sup> Eurogentec products are sold for research or laboratory use only and are not to be administrated to humans or used for medical diagnostics.

# ⚠ ROX™ Adjustment before first use

## IMPORTANT

Before first use, convert your Master Mix to the correct ROX™ level.

Table 1 shows how much ROX™ passive reference to add to Takyon® Etheris qPCR Master Mix, according to the thermocycler used.

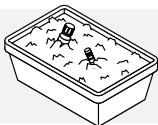
No ROX™ is needed for most thermocyclers not listed.

qPCR PLATFORM	1.5 ML MMX [ROX™ TO ADD]	7.5 ML MMX [ROX™ TO ADD]
Mx3000P®, 3005P, 4000	12 µL	60 µL
ABI Prism® 7500, FAST 7500, ViiA7™, QuantStudio™	4.2 µL	21 µL
ABI Prism® 5700, 7000, 7300, 7700, 7900, FAST 7900, ABI Step One, One Plus MasterCycler® ep realplex I & II (rev. <= 2.1)	60 µL	300 µL

Table 1 : ROX™ addition according to qPCR Platform.

## Recommended protocol

❶ Thaw all required reagents completely and put them on ice.

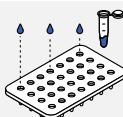


❷ Prepare the REACTION MIX in excess to correct for dispensing losses (e.g. a 100-reactions mix for 96-reactions). Add all components together, except for the template. Mix thoroughly by pipetting or inversion. Spin down.

REACTION MIX COMPONENTS	VOLUME / REACTION	FINAL CONCENTRATION
Takyon® MMx	5 µL	1x
Forward primer	2 µL	50-300 nM[3]
Reverse primer	2 µL	50-300 nM[3]
RNAse-free Water	x µL[2]	(volume is 20 µL minus all other components)[4]
<b>Total Mix/Reaction</b>	<b>20 µL</b>	

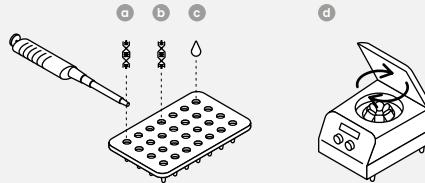
Table 2 : Reaction mix preparation.

❸ Add the REACTION MIX to individual reaction vials.



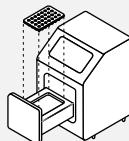
❹ Pipette into your qPCR plate/tubes either[4]:

a 2.5 µL of template cDNA / DNA   b 2.5 µL of positive control  
c 2.5 µL of water / buffer for negative control



d Gently mix on a magnetic stirrer and centrifuge to avoid bubbles.

❺ Program the Real-Time thermocycler using the following recommended parameters:



Takyon® Etheris produces consistent and sensitive results under FAST[5] cycling conditions.

	T°C	TIME
Takyon® activation	95°C	3 min
40 CYCLES[5]		
Denaturation[6]	95°C	3 -10 sec
Annealing / extension[7]	60°C	20 - 60 sec

Table 3 : Cycling conditions.

Trademarks : ABI Prism® 5700, 7000, 7500, 7700 and 7900 are registered trademarks of Applied Biosystems, Inc. | MasterCycler® is a registered trademark of Eppendorf, Inc. | Mx3000P®, Mx3005P® and Mx4000® are registered trademark of Stratagene | ROX™ is a trademark of Applied Biosystems Corp. | SYBR® Green is a registered trademark of Life Technologies Corp. | Takyon® is a registered trademark of Kaneka Eurogenetic S.A.

[2] Adjust water so total volume is 20 µL after adding all components. [3] Start with primers at 100 nM; optimize if needed. [4] Mix 17.5 µL reaction mix with 2.5 µL DNA for a 20 µL final volume; adjust if required. [5] Use FAST cycling only on FAST-block cyclers; prefer amplicons <120 bp [6] For complex templates (e.g., plant/genomic DNA), increase denaturation time. [7] Set annealing temperature according to primer Tm. For longer/difficult templates, extend annealing/extension to 40 s. Some FAST cyclers allow shorter denaturation/annealing for quicker qPCR; if amplification is poor, lengthen extension in 5 s steps.